

Amplite™ Colorimetric NADP/NADPH Ratio Assay Kit

Catalog number: 15274
Unit size: 250 Tests

| Component | Storage | Amount |
|--|--|--------------------------------|
| Component A: NADP/NADPH Recycling Enzyme Mix | Freeze (< -15 °C), Minimize light exposure | 2 bottles (lyophilized powder) |
| Component B-I: NADPH Probe | Freeze (< -15 °C), Minimize light exposure | 1 bottle (4 mL) |
| Component B-II: NADPH Probe Buffer | Freeze (< -15 °C), Minimize light exposure | 1 bottle (16 mL) |
| Component C: NADPH Standard | Freeze (< -15 °C), Minimize light exposure | 1 vial (167 µg) |
| Component D: NADP Extraction Solution | Freeze (< -15 °C), Minimize light exposure | 1 bottle (10 mL) |
| Component E: Neutralization Solution | Freeze (< -15 °C), Minimize light exposure | 1 bottle (10 mL) |
| Component F: Extraction Control Solution | Freeze (< -15 °C), Minimize light exposure | 1 bottle (10 mL) |
| Component G: Lysis Buffer | Freeze (< -15 °C), Minimize light exposure | 1 bottle (10 mL) |

OVERVIEW

Nicotinamide adenine dinucleotide (NAD⁺) and nicotinamide adenine dinucleotide phosphate (NADP⁺) are two important cofactors found in cells. NADH is the reduced form of NAD⁺, and NAD⁺ is the oxidized form of NADH. It forms NADP with the addition of a phosphate group to the 2' position of the adenylyl nucleotide through an ester linkage. NADP is used in anabolic biological reactions, such as fatty acid and nucleic acid synthesis, which require NADPH as a reducing agent. The traditional NAD/NADH and NADP/NADPH assays are done by monitoring of NADH or NADPH absorption at 340 nm. This method suffers low sensitivity and high interference since the assay is done in the UV range that requires expensive quartz microplate. Our Amplite™ NADP/NADPH Ratio Assay Kit provides a convenient method for sensitive detection of NADP, NADPH and their ratio. The NADPH probe is a chromogenic sensor that has its maximum absorbance at ~460 nm upon NADH reduction. The absorbance increase at ~460 nm is directly proportional to the concentration of NADPH in the solution. The NADPH probe can recognize NADPH in an enzyme-free reaction, and the signal can be easily read by an absorbance microplate reader at ~460 nm. The Amplite™ Colorimetric NADPH Assay Kit provides a sensitive assay to detect as little as 3 µM NADPH in a 100 µL assay volume. The assay can be performed in a convenient 96-well or 384-well microtiter-plate format.

AT A GLANCE

Protocol Summary

1. Prepare 25 µL of NADPH standards and/or test samples
2. Add 25 µL of NADP Extraction Solution
3. Incubate at 37 °C for 15 minutes
4. Add 25 µL of Neutralization Solution
5. Add 75 µL of NADP/NADPH working solution
6. Incubate at RT for 15 minutes to 2 hours
7. Monitor Absorbance at 460 nm

Important

1. It is highly recommended to incubate the cells with Lysis Buffer (Component G) at 37 °C and use the supernatant for the experiment.
2. Thaw one of each kit component at room temperature before starting the experiment.

KEY PARAMETERS

Absorbance microplate reader

Absorbance 460 nm
Recommended plate Clear bottom

CELL PREPARATION

For guidelines on cell sample preparation, please visit <https://www.aatbio.com/resources/guides/cell-sample-preparation.html>

PREPARATION OF STOCK SOLUTIONS

Unless otherwise noted, all unused stock solutions should be divided into single-use aliquots and stored at -20 °C after preparation. Avoid repeated freeze-thaw cycles.

NADPH standard solution (1 mM)

Add 200 µL of PBS buffer into the vial of NADPH standard (Component C) to have 1 mM (1 nmol/µL) NADPH stock solution.

PREPARATION OF STANDARD SOLUTION

For convenience, use the Serial Dilution Planner:
<https://www.aatbio.com/tools/serial-dilution/15274>

NADPH standard

Add 10 µL of 1 mM NADPH stock solution into 490 µL PBS buffer (pH 7.4) to generate 20 µM (20 pmols/µL) NADPH standard solution. Then take the 20 µM NADPH standard solution and perform 1:2 serial dilutions to get remaining serially diluted NADPH standards (NS7 - NS1). Note: Diluted NADPH standard solution is unstable, and should be used within 4 hours.

PREPARATION OF WORKING SOLUTION

1. Add 8 mL of NADPH Probe buffer (Component B-II) to the bottle of NADP/NADPH Recycling Enzyme Mixture (Component A), and mix well.
2. Add 2 mL NADPH Probe (Component B-I) into the same bottle (from Step 1) and mix well.

Note This NADP/NADPH working solution is enough for 125-200 assays. The working solution is not stable, use it promptly and avoid direct exposure to light.

SAMPLE EXPERIMENTAL PROTOCOL

TOTAL NADP/NADPH assay (avail. 400 assays/kit):

Table 1. Layout of NADPH standards and test samples in a white/clear bottom 96-well microplate. NS= NADPH Standards (NS1-NS7, 0.156 to 10 µM), BL=Blank Control, TS=Test Samples.

| | | | |
|-----|-----|-----|-----|
| BL | BL | TS | TS |
| NS1 | NS1 | ... | ... |
| NS2 | NS2 | ... | ... |
| NS3 | NS3 | | |
| NS4 | NS4 | | |
| NS5 | NS5 | | |
| NS6 | NS6 | | |
| NS7 | NS7 | | |

Table 2. Reagent composition for each well.

Note High concentration of NADPH (e.g., >30 μ M, final concentration) will cause saturated signal and make the calibration curve non-linear.

| Well | Volume | Reagent |
|-----------|------------|--|
| NS1 - NS7 | 50 μ L | Serial Dilutions (0.156 to 10 μ M) |
| BL | 50 μ L | PBS |
| TS | 50 μ L | Test Sample |

1. Prepare NADPH standards (NS), blank controls (BL), and test samples (TS) according to the layout provided in Tables 1 and 2.

Note Prepare cells or tissue samples as desired. Lysis Buffer (Component G) can be used for lysing the cells for convenience.

Note It is highly recommended to incubate the cells with Lysis Buffer (Component G) at 37 ° C and use the supernatant for the experiment.

2. Add 50 μ L of NADP/NADPH working solution into each well of NADPH standard, blank control, and test samples to make the total NADP/NADPH assay volume of 100 μ L/well.
3. Incubate the reaction at room temperature for 15 minutes to 2 hours, protected from light. (We used 1 hour incubation time in data shown)
4. Monitor the absorbance increase with an absorbance plate reader at 460 nm.

NADP/NADPH RATIO assay (avail. 250 assays/kit):

Table 3. Layout of NADPH standards and test samples in a white/clear 96-well microplate. NS= NADP/NADPH Standards (NS1 - NS7, 0.156 to 10 μ M); BL=Blank Control; TS=Test Samples; TS (NADP) = Test Samples treated with NADP Extraction Solution (Component D) for 15 minutes, then neutralized by Neutralization Solution (Component E).

| BL | BL | TS | TS | TS (NADP) | TS (NADP) |
|-----|-----|-----|-----|-----------|-----------|
| NS1 | NS1 | ... | ... | | |
| NS2 | NS2 | ... | ... | | |
| NS3 | NS3 | | | | |
| NS4 | NS4 | | | | |
| NS5 | NS5 | | | | |
| NS6 | NS6 | | | | |
| NS7 | NS7 | | | | |

Table 4. Reagent compositions for each well.

Note High concentration of NADPH (e.g., >30 μ M, final concentration) will cause saturated signal and make the calibration curve non-linear.

| NADPH Standard | Blank Control | Test Sample (NADP/NADPH) | Test Sample (NADP Extract) |
|---------------------------------|---------------------------------|---------------------------------|---------------------------------|
| Serial Dilutions: 25 μ L | PBS: 25 μ L | Test Sample: 25 μ L | Test Sample: 25 μ L |
| Component F: 25 μ L | Component F: 25 μ L | Component F: 25 μ L | Component D: 25 μ L |
| Incubate at 37°C for 15 minutes | Incubate at 37°C for 15 minutes | Incubate at 37°C for 15 minutes | Incubate at 37°C for 15 minutes |
| Component F: 25 μ L | Component F: 25 μ L | Component F: 25 μ L | Component E: 25 μ L |
| Total: 75 μ L | Total: 75 μ L | Total: 75 μ L | Total: 75 μ L |

1. Refer to Tables 3 & 4 for compositions of each well.
 - a. **For NADP Extraction (NADP amount):** Add 25 μ L of NADP Extraction Solution (Component D) into the wells of NADP/NADPH containing test samples. Incubate at 37 ° C for 10 to 15 minutes, then add 25 μ L of Neutralization Solution (Component E) to neutralize the NADP extracts as described in Tables 3 & 4.
 - b. **For Total NADP and NADPH (Total amount):** Add 25 μ L of NADP/NADPH Control Solution (Component F) into the wells of NADPH standards and NADP/NADPH containing test samples. Incubate at 37°C for 10 to 15 minutes, and then add 25 μ L of Extraction Control Solution (Component F) as described in Tables 3 and 4.

Note Prepare cells or tissue samples as desired. Lysis Buffer (Component G) can be used for lysing the cells for convenience.

2. Add 75 μ L of NADP/NADPH working solution into each well of NADPH standard, blank control, and test samples (NADP/NADPH), and test sample (NADP Extract) to make the total assay volume of 150 μ L/well.
3. Incubate the reaction at room temperature for 15 minutes to 2 hours, protected from light.
4. Monitor the absorbance increase with an absorbance plate reader at 460 nm.

EXAMPLE DATA ANALYSIS AND FIGURES

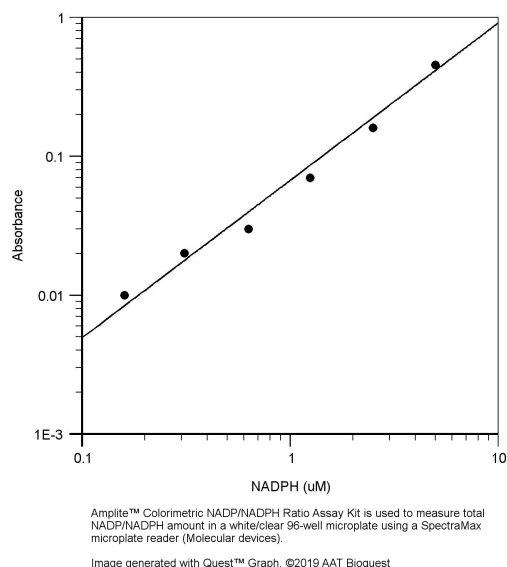


Figure 1. Amplite™ Colorimetric NADP/NADPH Ratio Assay Kit is used to measure total NADP/NADPH amount in a white/clear 96-well microplate using a SpectraMax microplate reader (Molecular devices).

DISCLAIMER

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