

INTENDED USE

This mouse albumin antigen assay is intended for the quantitative determination of total albumin in mouse plasma, serum, urine & other biological fluids. **For research use only.**

BACKGROUND

Albumin is a water-soluble protein with considerable structural stability which makes up 60% of the total protein of plasma. It functions as a carrier of hormones, enzymes, fatty acids, metal ions, and medicinal products.

ASSAY PRINCIPLE

Mouse albumin will bind to the affinity purified capture antibody coated on the microtiter plate. After appropriate washing steps, peroxidase labeled polyclonal anti-mouse albumin antibody binds to the captured protein. Excess antibody is washed away and TMB substrate is used for color development at 450nm. A standard calibration curve is prepared along with the samples to be measured using dilutions of mouse albumin. Color development is proportional to the concentration of albumin in the samples.

REAGENTS PROVIDED

- **96-well antibody coated microtiter strip plate** (removable wells 8x12) containing anti-mouse albumin antibody, blocked and dried.
- **10X Wash buffer:** 1 bottle of 50ml
- **5X Diluent:** 1 bottle of 50ml
- **Mouse albumin standard:** 1 vial lyophilized standard
- **Anti-mouse albumin primary antibody:** 1 vial concentrated polyclonal antibody
- **TMB substrate solution:** 1 bottle of 10ml solution

STORAGE AND STABILITY

Store all kit components at 4°C upon arrival. Return any unused microplate strips to the plate pouch with desiccant. Reconstituted standard may be stored at -80°C for later use. Do not freeze-thaw the standard more than once. Store all other unused kit components at 4°C. This kit should not be used beyond the expiration date.

OTHER REAGENTS AND SUPPLIES REQUIRED

- Microtiter plate shaker capable of 300 rpm uniform horizontally circular movement
- Manifold dispenser/aspirator or automated microplate washer
- Microplate reader capable of measuring absorbance at 450 nm
- Pipettes and Pipette tips
- Deionized or distilled water
- Polypropylene tubes for dilution of standard
- Paper towels or laboratory wipes
- 1N H₂SO₄ or 1N HCl

PRECAUTIONS

- **FOR LABORATORY RESEARCH USE ONLY. NOT FOR DIAGNOSTIC USE.**
- Do not mix any reagents or components of this kit with any reagents or components of any other kit. This kit is designed to work properly as provided.
- Always pour peroxidase substrate out of the bottle into a clean test tube. Do not pipette out of the bottle as contamination could result.
- Keep plate covered except when adding reagents, washing, or reading.
- **DO NOT** pipette reagents by mouth and avoid contact of reagents and specimens with skin.
- **DO NOT** smoke, drink, or eat in areas where specimens or reagents are being handled.

PREPARATION OF REAGENTS

- **1X Diluent:** 5X Diluent may contain precipitate. Warm to redissolve before use. Dilute 50ml of 5X diluent concentrate with 200ml of deionized water.
- **1X Wash buffer:** Dilute 50ml of 10X wash buffer concentrate with 450ml of deionized water.

SAMPLE COLLECTION

Collect plasma using EDTA or citrate as an anticoagulant. Centrifuge for 15 minutes at 1000xg within 30 minutes of collection. Assay immediately or aliquot and store at ≤ -20°C. Avoid repeated freeze-thaw cycles. Urine samples may be affected by freeze-thaw cycles or centrifugation.

ASSAY PROCEDURE

Perform assay at room temperature. Vigorously shake plate (300rpm) at each step of the assay.

Preparation of Standard

Reconstitute standard by adding 1ml of diluent directly to the vial and agitate gently to completely dissolve contents. This will result in a 1000ng/ml standard solution.

Dilution table for preparation of mouse albumin standard:

Albumin concentration (ng/ml)	Dilutions
1000	Straight from the vial
500	500µl Diluent + 500µl (1000ng/ml)
200	600µl Diluent + 400µl (500ng/ml)
100	500µl Diluent + 500µl (200ng/ml)
50	500µl Diluent + 500µl (100ng/ml)
20	600µl Diluent + 400µl (50ng/ml)
10	500µl Diluent + 500µl (20ng/ml)
5	500µl Diluent + 500µl (10ng/ml)
2	600µl Diluent + 400µl (5ng/ml)
1	500µl Diluent + 500µl (2ng/ml)
0	500µl Diluent Zero point to determine background

NOTE: DILUTIONS FOR THE STANDARD CURVE AND ZERO STANDARD MUST BE MADE AND APPLIED TO THE PLATE IMMEDIATELY.

Standard and Unknown Addition

Remove microtiter plate from bag and add 100µl albumin standards (in duplicate) and unknowns to wells. Carefully record position of standards and unknowns. Shake plate at 300rpm for 30 minutes. Wash wells three times with 300µl wash buffer. Remove excess wash by gently tapping plate on paper towel or kimwipe.

NOTE: The assay measures albumin antigen in the 1-1000 ng/ml range. If the unknown is thought to have high albumin levels, dilutions may be made in diluent. A 1:1,000,000-1:4,000,000 dilution for normal mouse plasma or a 1:1,000 dilution for normal mouse urine is suggested for best results.

Primary Antibody Addition

Briefly centrifuge vial before opening. Dilute 4µl of conjugated primary antibody in 10ml of diluent and add 100µl to all wells. Shake plate at 300rpm for 30 minutes. Wash wells three times with 300µl wash buffer. Remove excess wash by gently tapping plate on paper towel or kimwipe.

Substrate Incubation

Add 100µl TMB substrate to all wells and shake plate for 2-10 minutes. Substrate will change from colorless to different strengths of blue. Quench reaction by adding 50µl of 1N H₂SO₄ or HCl stop solution to all wells when samples are visually in the same range as the standards. Add stop solution to wells in the same order as substrate upon which color will change from blue to yellow. Mix thoroughly by gently shaking the plate.

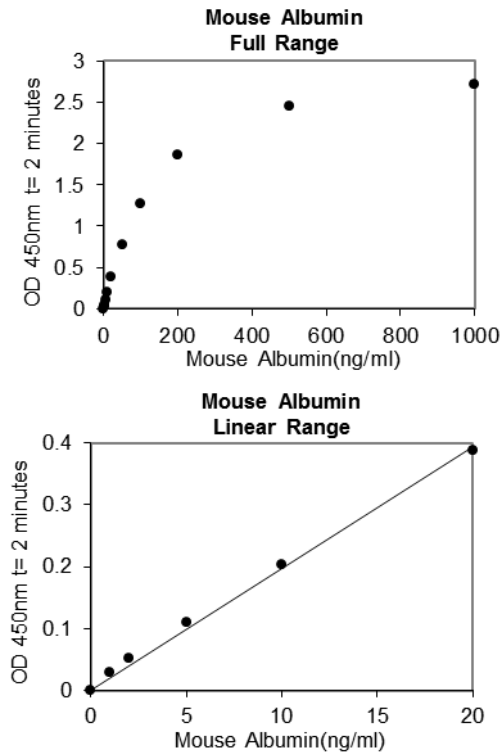
Measurement

Set the absorbance at 450nm in a microtiter plate spectrophotometer. Measure the absorbance in all wells at 450nm. Subtract zero point from all standards and unknowns to determine corrected absorbance (A₄₅₀).

Calculation of Results

Plot A₄₅₀ against the amount of albumin in the standards. Fit a straight line through the linear points of the standard curve using a linear fit procedure if unknowns appear on the linear portion of the standard curve. Alternatively, create a standard curve by analyzing the data using a software program capable of generating a four parameter logistic (4PL) curve fit. The amount of albumin in the unknowns can be determined from this curve. If samples have been diluted, the calculated concentration must be multiplied by the dilution factor.

A typical standard curve (EXAMPLE ONLY):



EXPECTED VALUES

Albumin is present in normal mouse serum at a concentration of 20 mg/ml in Balb/C, 27 mg/ml in C57BL6, and 29 mg/ml in CD1 strains [1].

PERFORMANCE CHARACTERISTICS

Sensitivity: The minimum detectable dose (MDD) was determined by adding two standard deviations to the mean optical density value of twenty zero standard replicates (range OD₄₅₀: 0.042-0.05) and calculating the corresponding concentration. The MDD was 0.2 ng/ml.

Intra-assay Precision: Three samples of known concentration were tested twenty times on one plate to assess intra-assay precision.

Sample	1	2	3
n	20	20	20
Mean (ng/ml)	20.6	81.3	185
Standard Deviation	1.30	3.26	11.1
CV (%)	6.31	4.01	5.98

Inter-assay Precision: These studies are currently in progress. Please contact us for more information.

Recovery: These studies are currently in progress. Please contact us for more information.

Linearity: To assess the linearity of the assay, samples of blocking buffer spiked with high concentrations of antigen were serially diluted to produce samples with values within the dynamic range of the assay.

Sample	1:2	1:4	1:8	1:16
n	4	4	4	4
Average % of Expected	99	100	96	93
Range	96-101%	94-104%	92-101%	78-107%

Specificity: Pooled normal plasma from rat was assayed and slight cross-reactivity was observed. Pooled normal plasma from rabbit, dog, pig, sheep, cyno monkey, and human was assayed and no significant cross-reactivity was observed.

Sample Values: Samples were evaluated for the presence of the antigen at varying dilutions.

Sample Type	Dilution	Mean (µg/mL)
CD-1 Citrate Plasma	1:1,000,000	29,000
BALB/c Urine	1:1,000	33
Nude Mouse Urine	1:1,000	19

DISCLAIMER

This information is believed to be correct but does not claim to be all-inclusive and shall be used only as a guide. The supplier of this kit shall not be held liable for any damage resulting from handling of or contact with the above product.

REFERENCES

1.Zaias J, *et al.*: J Am Assoc Lab Anim Sci. 2009, 48(4):387-390.

SAMPLE INSERT
Refer to kit box for lot specific instructions

Example of ELISA Plate Layout

96 Well Plate: 22 Standard wells, 74 Sample wells

	1	2	3	4	5	6	7	8	9	10	11	12
A	0	1 ng/ml	2 ng/ml	5 ng/ml	10 ng/ml	20 ng/ml	50 ng/ml	100 ng/ml	200 ng/ml	500 ng/ml	1000 ng/ml	
B	0	1 ng/ml	2 ng/ml	5 ng/ml	10 ng/ml	20 ng/ml	50 ng/ml	100 ng/ml	200 ng/ml	500 ng/ml	1000 ng/ml	
C												
D												
E												
F												
G												
H												