

# Fura-8FF™. AM

Catalog number: 20620 Unit size: 10x50 ug

Component	Storage	Amount
Fura-8FF™, AM	Freeze (< -15 °C), Minimize light exposure	10x50 ug

## **OVERVIEW**

The cell-permeant Fura-8FF AM is an analog of Fura-8 AM with much lower calcium binding affinity, Kd  $\sim$ 10  $\mu$ M. Fura-8FF has its emission shifted into longer visible wavelength that is compatible with the common filter sets. Fura-8FF  $^{TM}$  AM is more sensitive to calcium than Fura-2FF AM with higher signal/background ratio than that of Fura-2FF AM. e., calculating the excitation intensity ratios at 354 nm and 415 nm by monitoring emission intensity at 530 nm.

## **KEY PARAMETERS**

#### Fluorescence microscope

Excitation Fura 2 filter set
Emission Fura 2 filter set
Recommended plate Black wall/clear bottom

#### Fluorescence microplate reader

 Excitation
 355, 415

 Emission
 530

 Cutoff
 475

Recommended plate Black wall/clear bottom

Instrument specification(s)

Bottom read mode/Programmable liquid

handling

### PREPARATION OF STOCK SOLUTIONS

Unless otherwise noted, all unused stock solutions should be divided into single-use aliquots and stored at -20 °C after preparation. Avoid repeated freeze-thaw cycles.

### Fura-8FF™ AM Stock Solution

Prepare a 2 to 5 mM stock solution of Fura-8FF™ AM in high-quality, anhydrous DMSO.

## PREPARATION OF WORKING SOLUTION

## Fura-8FF™ AM Working Solution

On the day of the experiment, either dissolve Fura-8FF<sup>TM</sup> AM in DMSO or thaw an aliquot of the indicator stock solution to room temperature. Prepare a dye working solution of 2 to 20  $\mu$ M in a buffer of your choice (e.g., Hanks and Hepes buffer) with 0.04% Pluronic® F-127. For most cell lines, Fura-8FF<sup>TM</sup> AM at a final concentration of 4-5  $\mu$ M is recommended. The exact concentration of indicators required for cell loading must be determined empirically.

**Note** The nonionic detergent Pluronic® F-127 is sometimes used to increase the aqueous solubility of Fura-8FF™ AM. A variety of Pluronic® F-127 solutions can be purchased from AAT Bioquest.

**Note** If your cells contain organic anion-transporters, probenecid (1-2 mM) may be added to the dye working solution (final in well concentration will be 0.5-1 mM) to reduce leakage of the de-esterified indicators. A variety of ReadiUse<sup>™</sup> probenecid products, including water-soluble, sodium salt, and stabilized solution, can be purchased from AAT Bioquest.

# SAMPLE EXPERIMENTAL PROTOCOL

Following is our recommended protocol for loading AM esters into live cells. This protocol only provides a guideline and should be modified according to your specific needs.

- 1. Prepare cells in growth medium overnight.
- On the next day, add 1X Fura-8FF™ AM working solution into your cell plate.

**Note** If your compound(s) interfere with the serum, replace the growth medium with fresh HHBS buffer before dye-loading.

 Incubate the dye-loaded plate in a cell incubator at 37 °C for 30 to 60 minutes.

**Note** Incubating the dye for longer than 1 hour can improve signal intensities in certain cell lines.

- Replace the dye working solution with HHBS or buffer of your choice (containing an anion transporter inhibitor, such as 1 mM probenecid, if applicable) to remove any excess probes.
- 5. Add the stimulant as desired and simultaneously measure fluorescence using either a fluorescence microscope equipped with a Fura 2 filter set or a fluorescence plate reader containing a programmable liquid handling system such as a FlexStation, at Ex/Em <sub>1</sub> = 355/530 nm cutoff 475 nm and Ex/Em <sub>2</sub> = 415/530 nm cutoff 475 nm.

## **EXAMPLE DATA ANALYSIS AND FIGURES**

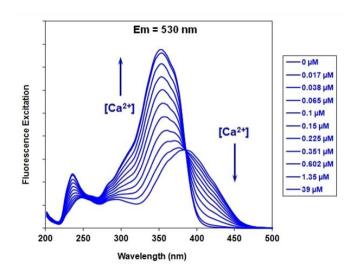


Figure 1. Fluorescence excitation spectra of Fura- $8^{TM}$  in the presence of 0 to 39  $\mu M$  free Ca2+.

## **DISCLAIMER**

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