

## Rhod-FF, AM

 Catalog number: 21077, 21078  
 Unit size: 1 mg, 10x50 ug

Component	Storage	Amount (Cat No. 21077)	Amount (Cat No. 21078)
Rhod-FF, AM	Freeze (< -15 °C), Minimize light exposure	1 vial (1 mg)	10x50 ug

### OVERVIEW

Calcium measurement is critical for numerous biological investigations. Fluorescent probes that show spectral responses upon binding Ca<sup>2+</sup> have enabled researchers to investigate changes in intracellular free Ca<sup>2+</sup> concentrations by using fluorescence microscopy, flow cytometry, fluorescence spectroscopy and fluorescence microplate readers. Rhod-FF AM is cell-permeable, and generates Rhod-FF upon esterase hydrolysis in cells. Rhod-FF has a lower binding affinity for Ca<sup>2+</sup> and is suitable for Ca<sup>2+</sup> measurements from 10 to 200 uM. Like the parent Rhod-2 indicator, Rhod-FF is essentially nonfluorescent in the absence of divalent cations and exhibits strong fluorescence enhancement with no spectral shift upon binding Ca<sup>2+</sup>.

### KEY PARAMETERS

#### Fluorescence microscope

Excitation	TRITC filter set
Emission	TRITC filter set
Recommended plate	Black wall/clear bottom

#### Fluorescence microplate reader

Excitation	540
Emission	590
Cutoff	570
Recommended plate	Black wall/clear bottom
Instrument specification(s)	Bottom read mode/Programmable liquid handling

### PREPARATION OF STOCK SOLUTIONS

Unless otherwise noted, all unused stock solutions should be divided into single-use aliquots and stored at -20 °C after preparation. Avoid repeated freeze-thaw cycles.

#### Rhod-FF AM Stock Solution

Prepare a 2 to 5 mM stock solution of Rhod-FF AM in high-quality, anhydrous DMSO.

### PREPARATION OF WORKING SOLUTION

#### Rhod-FF AM Working Solution

On the day of the experiment, either dissolve Rhod-FF AM in DMSO or thaw an aliquot of the indicator stock solution to room temperature. Prepare a dye working solution of 2 to 20 μM in a buffer of your choice (e.g., Hanks and Hepes buffer) with 0.04% Pluronic® F-127. For most cell lines, Rhod-FF AM at a final concentration of 4-5 μM is recommended. The exact concentration of indicators required for cell loading must be determined empirically.

**Note** The nonionic detergent Pluronic® F-127 is sometimes used to increase the aqueous solubility of Rhod-FF AM. A variety of Pluronic® F-127 solutions can be purchased from AAT Bioquest.

**Note** If your cells contain organic anion-transporters, probenecid (1-2 mM) may be added to the dye working solution (final in well concentration will be 0.5-1 mM) to reduce leakage of the de-esterified indicators. A variety of ReadiUse™ probenecid products, including water-soluble, sodium salt, and stabilized solution, can be purchased from AAT Bioquest.

### SAMPLE EXPERIMENTAL PROTOCOL

Following is our recommended protocol for loading AM esters into live cells. This protocol only provides a guideline and should be modified according to your specific needs.

1. Prepare cells in growth medium overnight.
2. On the next day, add 1X Rhod-FF AM working solution into your cell plate.

**Note** If your compound(s) interfere with the serum, replace the growth medium with fresh HHBS buffer before dye-loading.

3. Incubate the dye-loaded plate in a cell incubator at 37 °C for 30 to 60 minutes.

**Note** Incubating the dye for longer than 1 hour can improve signal intensities in certain cell lines.

4. Replace the dye working solution with HHBS or buffer of your choice (containing an anion transporter inhibitor, such as 1 mM probenecid, if applicable) to remove any excess probes.
5. Add the stimulant as desired and simultaneously measure fluorescence using either a fluorescence microscope equipped with a TRITC filter set or a fluorescence plate reader containing a programmable liquid handling system such as an FDSS, FLIPR, or FlexStation, at Ex/Em = 540/590 nm cutoff 570 nm.

### EXAMPLE DATA ANALYSIS AND FIGURES

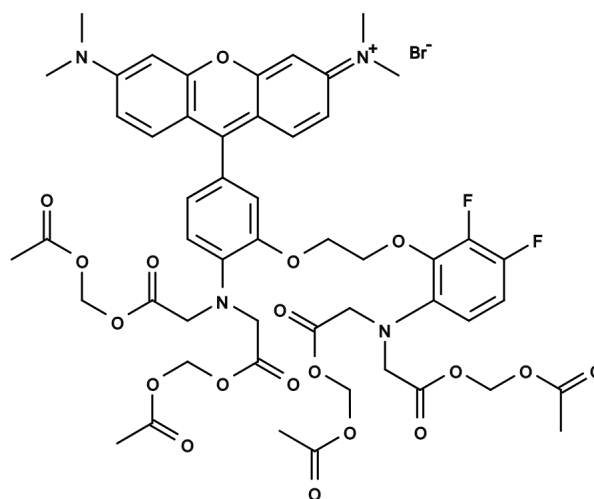


Figure 1. Chemical structure for Rhod-FF, AM

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