

## Buccutite™ FOL-Dye 650

Catalog number: 5372

Unit size: 2 umoles

| Component              | Storage                                   | Amount   |
|------------------------|---|----------|
| Buccutite™ FOL-Dye 650 | Freeze (<-15 °C), Minimize light exposure | 2 umoles |

### OVERVIEW

Our Buccutite™ crosslinking technology provides the most convenient and effective crosslinking method to link two biomolecules with a high conjugation yield. Our method uses one pair of crosslinkers: Buccutite™ MTA and Buccutite™ FOL. MTA is added to one molecule, while FOL is added to another molecule. The cross-linking reaction is initiated by mixing Molecule-1-Buccutite™ MTA and Molecule-2-Buccutite™ FOL. This crosslinking reaction occurs under extremely mild and neutral conditions without any catalyst required. It is robust and efficient. Many of our customer have requested us to offer the stand-alone Buccutite™ MTA and Buccutite™ FOL reagents to expand the application of Buccutite™ crosslinking technology. This Buccutite™ FOL reagent is used to determine the number of FOL groups of the Molecule-2-Buccutite™ FOL. The number of FOL linkers provides an important parameter to optimize crosslinking process.

### PREPARATION OF STOCK SOLUTIONS

Unless otherwise noted, all unused stock solutions should be divided into single-use aliquots and stored at -20 °C after preparation. Avoid repeated freeze-thaw cycles.

*Buccutite™ FOL-Dye 650 stock solution (10 mM):*

Add 200 uL DMSO to Buccutite™ FOL-Dye 650 vial to prepare 10 mM stock solution.

**Note** The Buccutite™ FOL-Dye 650 stock solution should be stored at -20 °C after preparation and stable for 2 months if avoid repeated freeze-thaw cycles.

### SAMPLE EXPERIMENTAL PROTOCOL

#### FOL Sample Preparation

1. Use 100 ug FOL-modified sample (for example: antibody or other protein modified with MTA group, the MW should be above 15,000).
2. Adjust the volume to 100 uL with PBS.

#### Run FOL Assay

1. Add 10 uL 10 mM Buccutite™ FOL-Dye 650 stock solution to FOL sample solution.
2. Keep the reaction mixture at room temperature and rotate or shake it for 60 minutes.
3. Prepare spin column (Cat#60500) for sample purification.
4. Load the reaction mixture to a spin column with a clean collecting tube. After all the solution loaded to the column, add 10 uL PBS to the top and centrifuge the column for 5 minutes at 1,000 x g.
5. Collect the solution with a collecting tube.
6. Measure the absorption spectra with 0.5 mL Quartz Cuvette or Nanodrop.

**Note** Dilute the elution by 5 - 10 folds with PBS, measure the absorption spectrum from 800 nm to 250 nm, or only read the absorbance number at 280 nm and 654 nm.

7. Calculate FOL # (moles of FOL / mole of molecule) with the following equation.

$$\text{FOL \#} = (\text{A654} / 250000) / \{(\text{A280} - 0.09 \times \text{A654}) / \text{EC}\}$$

A280: absorbance of the elution at 280 nm  
 A654: absorbance of the elution at 654 nm  
 EC: Extinction Coefficient of the sample ( $\text{M}^{-1}\text{cm}^{-1}$ )

### EXAMPLE DATA ANALYSIS AND FIGURES

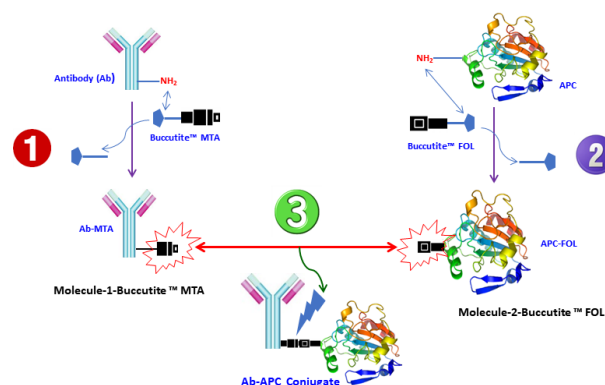
#### FOL Calculations:

**Sample:** GxM IgG-FOL, 100 ug in 100 uL PBS

Measure absorbance with a Nanodrop spectrophotometer,

A280 nm = 0.766,  
 A654 nm = 0.852,  
 CF280 = 0.09,  
 EC (Buccutite™ FOL-Dye 650) at 650 nm = 250,000  $\text{M}^{-1}\text{cm}^{-1}$   
 EC of IgG at 280 nm = 210,000  $\text{M}^{-1}\text{cm}^{-1}$

$$\text{FOL \# (moles of MTA per mole of IgG)} = (0.852 / 250000) / \{(0.766 - 0.09 \times 0.852) / 210000\} = 1.0$$



**Figure 1.** Buccutite™ crosslinking technology provides the most convenient and effective crosslinking method to link two biomolecules with a high conjugation yield. Our method uses one pair of crosslinkers: Buccutite™ MTA and Buccutite™ FOL. MTA is added to one molecule, while FOL is added to another molecule. The cross-linking reaction is initiated by mixing Molecule-1-Buccutite™ MTA and Molecule-2-Buccutite™ FOL. This crosslinking reaction occurs under extremely mild and neutral conditions without any catalyst required. It is robust and efficient.

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