

**Cell Meter™ BX520 fixable viability dye**

 Catalog number: 22510  
 Unit size: 200 Tests

| Component                               | Storage                                    | Amount    |
|---|--|-----------|
| Cell Meter™ BX520 fixable viability dye | Freeze (< -15 °C), Minimize light exposure | 200 tests |

**OVERVIEW**

Discrimination and exclusion of dead cells from live cells allows cleaner separation and identification of cell populations. Cell Meter™ fixable viability dyes are a large family of cell-impermeable fluorescent viability dyes that are optimized to match the major excitation lasers of common flow cytometers, such as 350, 405, 488, 633 and 647 nm. These dyes are impermeant to live cells but permeant to cells with compromised membranes. They irreversibly react with amine- and thiol-containing proteins and other cellular components. Since dead or fixed cells with a compromised membrane more readily react with Cell Meter™ fixable cell stains, thus stain brighter than live cells with an intact membrane, these dyes can be used to assess live vs. dead status of mammalian cells. There are a few factors to be considered when using these dyes, e.g., the titration of each dye to ensure that live cells have minimal to no staining. Cell Meter™ BX520 fixable cell stain is optimized to be excited with the blue laser at 488 nm with emission at 520 nm (FITC channel). Compared to other commercially similar viability dyes, this fixable viability dye is much more robust and stable.

**AT A GLANCE**
**Protocol summary**

1. Prepare samples in HHBS (0.5 mL/assay)
2. Add Cell Meter™ BX520 to the cell suspension
3. Stain the cells at room temperature for 20 - 60 minutes
4. Wash the cells
5. Fix the cells (optional)
6. Examine the sample with flow cytometer using 530/30 nm filter (FITC channel) and/or fluorescence microscope using FITC filter set

**Important**

Thaw at room temperature before starting the experiment.

**KEY PARAMETERS**
**Flow cytometer**

|                             |                  |
|-----------------------------|------------------|
| Excitation                  | 488 nm laser     |
| Emission                    | 530/30 nm filter |
| Instrument specification(s) | FITC filter set  |

**Fluorescence microscope**

|                   |                         |
|-------------------|-------------------------|
| Excitation        | FITC filter set         |
| Emission          | FITC filter set         |
| Recommended plate | Black wall/clear bottom |

**PREPARATION OF STOCK SOLUTIONS**

Unless otherwise noted, all unused stock solutions should be divided into single-use aliquots and stored at -20 °C after preparation. Avoid repeated freeze-thaw cycles.

**Cell Meter™ BX520 stock solution (500X)**

Add 200 µL of DMSO into the Cell Meter™ BX520 vial to make 500X stock solution.

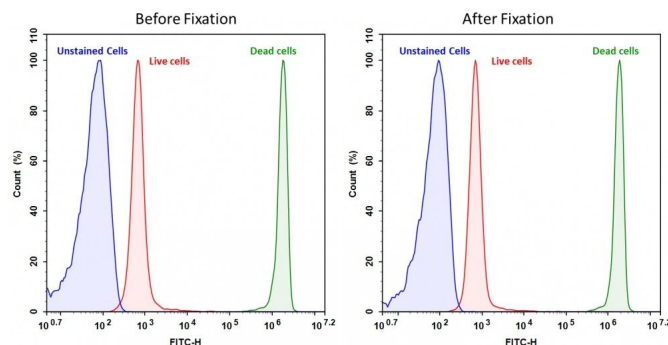
**Note** The unused Cell Meter™ BX520 stock solution should be divided into single use aliquots and stored at -20 °C. Avoid repeated freeze/thaw cycles.

**SAMPLE EXPERIMENTAL PROTOCOL**

1. Treat cells as desired.
2. Wash cells once with HHBS or the azide- and serum/protein-free buffer of your choice.
3. Resuspend cells at 5 - 10 × 10<sup>6</sup> /mL in HHBS or in the azide- and serum/protein-free buffer of your choice.
4. Add 1 µL of 500X Cell Meter™ BX520 stock solution to 0.5 mL of cells/assay and mix it well.
5. Incubate at room temperature, 5% CO<sub>2</sub> incubator for 20 - 60 minutes, protected from light.

**Note** The optimal stain concentrations and incubation time should be experimentally determined for different cell lines.

6. Wash cells twice and resuspend cells with HHBS or the buffer of your choice.
7. Fix cells as desired (optional).
8. Analyze cells with a flow cytometer using 530/30 nm filter (FITC channel) and/or fluorescence microscope using FITC filter set.

**EXAMPLE DATA ANALYSIS AND FIGURES**


**Figure 1.** Detection of Jurkat cell viability by Cell Meter™ fixable viability dye. Jurkat cells were treated and stained with Cell Meter™ BX520 (Cat#22510), and then fixed in 3.7% formaldehyde and analyzed by flow cytometry. The dead cell population (Green peak) is easily distinguished from the live cell population (Red peak) with FITC channel, and nearly identical results were obtained before and after fixation.

**DISCLAIMER**

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